

REVIEW

The prognostic power of gene mutations in thyroid cancer

Sara Ahmadi^{id} and Iñigo Landa^{id}

Division of Endocrinology, Thyroid Section, Department of Medicine, Brigham and Women's Hospital, Harvard Medical School, Boston, Massachusetts, USA

Correspondence should be addressed to S Ahmadi: sahmadi1@bwh.harvard.edu

Abstract

The introduction and generalization of next-generation sequencing techniques have significantly increased the identification of mutations in thyroid tumors from multiple patient cohorts. The understanding of the association between specific mutations and clinical outcomes is gradually leading to individualizing the care of patients with thyroid cancer. *BRAF*^{V600} is the most common mutation seen in thyroid cancer patients and unequivocally predicts malignancy, but when considered in isolation, it is not recommended to be used as an independent prognostic factor. Mutations in *RAS* are the second most common alterations in thyroid cancer but can be found in benign and malignant lesions. Rearrangements involving receptor tyrosine kinases, primarily *RET*, are found in a subset of thyroid tumors without mutations in either *BRAF* or *RAS*. The assessment of additional mutations is increasingly employed in thyroid cancer prognostication. The coexistence of *BRAF* with alterations in genes such as *PIK3CA*, *TERT* promoter, or *TP53* is associated with less favorable outcomes. Similar studies have also shown that additional oncogenic mutations in *RAS*-mutant thyroid carcinoma, such as those affecting the *EIF1AX* gene, likely predict a more aggressive clinicopathologic behavior. Overall, emerging evidence suggests that the co-occurrence of specific alterations in defined genes with *BRAF* or *RAS* mutations can become prognostic tools and useful predictors of thyroid tumor aggressiveness.

Keywords: thyroid cancer; genomic; prognosis

Introduction

Thyroid cancer is the most common malignancy of the endocrine system. Papillary thyroid carcinoma (PTC) is the most common thyroid carcinoma, accounting for almost 80% of cases; 12% of thyroid carcinomas have follicular (FTC) or oncocytic (OC; previously Hurthle cell) histology, <3% are poorly differentiated thyroid carcinoma (PDTC) and <2% are anaplastic thyroid carcinoma (ATC) (1). Differentiated thyroid cancers (DTC) encompass PTC, FTC, and OC, whereas PDTC and ATC are less differentiated and more aggressive. Our knowledge of the genomic landscape of thyroid cancer has dramatically improved over the last decade. The landmark study by The Cancer Genome Atlas (TCGA)

showed that PTC is dependent on the MAPK pathway activation via mutually exclusive mutations of *BRAF*, *RAS* or through gene rearrangements involving *RET* and other receptor tyrosine kinases (RTK) (2). Even though all these alterations activate MAPK signaling, *BRAF*-mutant, *RAS*-mutant, and *RET*-rearranged thyroid cancers have different behaviors. To date, several studies have aimed to establish genotype–phenotype correlations in patients with thyroid cancer, which has led to individualizing the care of patients with thyroid cancer, specifically patients with advanced thyroid cancer. This review article describes gene mutations in follicular cell-derived thyroid cancer with known prognostic power. Oncocytic

carcinoma has a different molecular foundation, including widespread chromosomal haploidization and mutations in the mitochondrial genome, none of which have yet been assessed for their prognostic value and hence not included in this review article.

BRAF mutations

BRAF mutations predominantly occur at codon V600 and are the most common alterations of PTC, as they are identified in around 60% of cases (2). Mutations in the *BRAF* gene drive thyroid cancer transformation via activation of the MAPK pathway. The *BRAF*^{V600E} mutation was initially associated with aggressive clinicopathologic features, including extrathyroidal extension, lymph node metastases (LNM), advanced stage, and poor clinical outcomes. In a meta-analysis of 19 studies on clinicopathologic features of papillary thyroid microcarcinoma (PTMC) with information on *BRAF* mutation, *BRAF*^{V600E} mutation was significantly associated with extrathyroidal extension, multifocality, LNM, and advanced stage (3). In a retrospective multicenter study (16 medical centers in eight countries) on 2099 PTC patients with a median follow-up of 36 months, *BRAF*^{V600E} mutation was associated with a higher risk of recurrence (47.71 vs 26.03 per 1000 person-years) with an unadjusted HR of 1.82. This remained significant after adjustment for patient age, sex, tumor size, extrathyroidal extension, LNM, and multifocality. Another retrospective multicenter study (11 centers in six countries) of 2,638 PTC patients with a median follow-up of 58 months showed synergism between LNM and *BRAF*^{V600E}; the disease recurrence rate was 10% vs 39.6% in non-LNM vs LNM patients, in the *BRAF*-positive group. The risk of mortality from cancer in this study was significantly higher in patients with LNM and *BRAF*^{V600E} (2.9% in patients only with LNM, 1.2% in patients with only *BRAF* vs 7.8% in patients with both *BRAF*^{V600E} and LNM; $P < 0.001$) (4).

However, other studies have questioned the value of the *BRAF*^{V600E} mutation as an independent prognostic factor (5, 6, 7, 8, 9). In a retrospective multicenter study (13 medical centers in 7 countries) of 1849 patients with PTC and a median follow-up of 33 months, *BRAF*^{V600E} mutation was associated with increased cancer-related mortality. However, when LNM, extrathyroidal extension, and distant metastases were included in the model, the association of *BRAF*^{V600E} with mortality was no longer statistically significant (10). In a retrospective study of 101 patients with PTMC (72 *BRAF*^{V600E} positive), there was no statistically significant association between *BRAF*^{V600E} and extrathyroidal extension, multifocality, and LNM (7). In another retrospective study of 631 PTC patients (38.4% *BRAF*^{V600E} positive), with an average of 83 months follow-up, *BRAF*^{V600E} mutation was not associated with extrathyroidal extension, lymph node, and distant metastases. Disease-free survival was not different in patients with vs without *BRAF* mutation

(6). In a retrospective study, 40 patients with PTMC with clinically significant lateral neck LNM and 71 patients with PTMC and no LNM were included. *BRAF* alterations were detected in 61% of the tumors. *BRAF*^{V600E} mutation did not predict clinically significant lateral neck LNM (8). Another retrospective study of 429 patients with PTC (9) showed no significant association between *BRAF*^{V600E} and more aggressive disease.

Although the association reported in multiple studies between *BRAF*^{V600E} and disease recurrence, the clinical application of *BRAF*^{V600E} alone as a prognostic marker is limited due to low specificity. Boucai *et al.*, in a study using TCGA data, showed that *BRAF* mutant tumors are heterogeneous. This study compared two subgroups of *BRAF*^{V600E} mutant tumors based on their thyroid differentiation score (TDS): those with a preserved expression of thyroid differentiation genes (*BRAF*-TDS^{hi}) vs their counterparts with decreased expression of thyroid differentiation genes (*BRAF*-TDS^{lo}). *BRAF*^{V600E} PTC tumors with downregulation of iodine metabolism genes (*BRAF*-TDS^{lo}) were larger ($P = 0.002$) and had higher T stage ($P = 0.002$) and more LNM ($P = 0.042$) and distant metastases ($P = 0.043$). There was no significant survival difference between the two groups ($P = 0.62$). Two out of 21 (9.5%) *BRAF*-TDS^{hi} patients had an incomplete response to therapy, whereas in the *BRAF*-TDS^{lo}, 28/76 (37%) patients had an incomplete response to therapy ($P < 0.01$) (11).

Overall, the 2015 American Thyroid Association guidelines recommended not using *BRAF*^{V600E} in isolation to guide the management of patients with low-risk thyroid carcinoma due to low positive predictive value (PPV) (1). However, the coexistence of *BRAF* with other oncogenic mutations, such as those in *PIK3CA*, *TERT* promoter, or *TP53*, is associated with less favorable outcomes, as discussed later in this review.

RAS mutations

Point mutations in the *RAS* genes (*NRAS*, *HRAS*, and *KRAS* isoforms) are DTC's second most common genetic alterations. Mutations in *RAS* genes drive thyroid cancer transformation via activation of the MAPK pathway but have lower oncogenic potential than *BRAF*^{V600E}. *BRAF*^{V600E}-mutant thyroid cancers (enriched in classic and tall cell subtypes) are more frequently invasive than *RAS*-mutant tumors and show a tropism to regional lymph nodes. *RAS*-mutant DTCs are typically encapsulated and rarely spread; when they do, e.g. in metastatic FTCs and PDTCs, they bypass lymph nodes and instead they more frequently affect bone and lung sites (12, 13). *RAS* mutations can be found in different thyroid tumors, including 10–20% of PTC, particularly in the follicular variant of PTC (fvPTC), 40–50% of FTC, and 20–40% of PDTC and ATC (14). The lower allelic frequency of *RAS* mutations in follicular adenomas than carcinomas might explain why they are found in benign

and malignant lesions (15). The use of *RAS* mutation as a molecular biomarker to diagnose thyroid cancer and to predict the prognosis is limited due to the heterogeneity described earlier. Instead, the identification of additional mutations in *RAS*-driven disease can inform prognosis. Some alterations, such as mutations in the *TERT* promoter and PI3K pathway effectors, occur across multiple DTCs, whereas others, such as mutations in *EIF1AX*, are strongly associated with *RAS*-mutant disease (16, 17). Studies assessing these additional alterations are summarized in the next section.

RET rearrangements

RET/PTC rearrangements generate fusion oncoproteins that reactivate RET. RET rearrangements are generated by the fusion of the catalytic domain of the tyrosine kinase receptor RET to the 5' terminal regions of heterologous genes. RET/PTC is associated explicitly with PTC and not with other types of thyroid cancer (18). Based on the TCGA study, the occurrence of RET/PTC in the PTC cohort is 6.8% (2).

Two groups of PTC harbor RET rearrangements: sporadic and radiation-induced. After the Chernobyl accident in 1986, there was a significant increase in the incidence of PTC in children, and the main driver was RET/PTC rearrangement (19, 20, 21, 22). Radiation-induced PTC with RET/PTC rearrangement has been reported to present at a more advanced T stage and have a higher probability of LNM. RET/PTC3 (NCOA4-RET) is predominant in radiation-induced PTC, and this rearrangement has been connected to the solid variant of PTC. RET/PTC1 (CCDC6-RET) is predominant in sporadic PTC and is strongly associated with the classic subtype (20, 23, 24). Radiation-induced PTC does not represent the general population with PTC and RET/PTC rearrangement. Studies on PTC with sporadic RET/PTC rearrangement have shown that these tumors are slow growing and do not usually progress toward more aggressive tumors like poorly differentiated thyroid carcinoma (25, 26). In pediatric patients, which are cohorts enriched for RTK fusions, RET/PTC-driven tumors have been associated with increased metastatic capacity and persistent disease compared to their *BRAF*- or *RAS*-mutant counterparts (27), whereas these differences are not observed in adult patients.

Additional mutations in thyroid cancer prognostication

Next-generation sequencing studies have shown that compared to DTC, PDTC and ATC display an extended set of mutations but typically harbor driver alterations in *BRAF*, *RAS*, or *RET* oncogenes (28, 29, 30). This suggests that PDTCs/ATCs can arise from PTCs/FTCs by accumulating key additional genetic abnormalities.

The idea of a continuum in disease progression can thus be used to inform genomics-driven prognostication in DTC patients. Mutations in genes for which the most robust studies exist are discussed below, and a subset of them is highlighted in Table 1.

TERT promoter mutations

Mutations in the proximal promoter of the *TERT* (telomerase reverse transcriptase) gene were initially discovered in melanomas (31, 32) and shortly after identified as frequent events in various tumor types, such as gliomas, hepatocellular, urothelial, and thyroid carcinomas (33, 34, 35, 36). *TERT* promoter mutations occur in the noncoding portion of the gene at either c.-124C>T or c.-146C>T, reactivating *TERT* transcription in thyroid cancer cells (36, 37, 38). *TERT* promoter mutations are relatively frequent events in DTCs, suggesting they could serve as biomarkers of disease progression. The fact that they occur rather early in tumor evolution implies that they likely enhance the proliferative properties of *BRAF*- or *RAS*-driven thyroid cancer clones.

The presence of a *TERT* promoter mutation in material from fine needle aspiration biopsies has been shown to offer a diagnostic specificity for malignancy close to 100%. It can help refine the assessment of thyroid nodules with an indeterminate cytology (39, 40, 41, 42, 43). Furthermore, multiple studies provided evidence of the prognostic value that the assessment of *TERT* mutations, either alone or, most efficiently, combined with *BRAF* or *RAS*, can provide for DTC patients. In a group of 51 PTC patients, Liu and colleagues showed that *TERT* promoter mutations were associated with metastatic disease and decreased survival (44). Melo *et al.* studied a cohort of 332 PTCs and observed that *TERT* mutations had an independent prognostic value: they were associated with increased rates of distant metastases, persistent disease, and disease-specific mortality (45). The cooperative effects that the co-occurrence of a *BRAF* or *RAS* mutation plus a *TERT* promoter mutation has in predicting poorer clinical outcomes have been widely studied, and it is now firmly established. In a cohort of 507 PTC patients, Xing and colleagues reported that coexisting *BRAF*^{V600E} and *TERT*^{-124C>T} mutations were associated with high-risk clinicopathologic characteristics, including increased recurrence rates and decreased recurrence-free survival (46). Several other studies in independent PTC cohorts showed associations of *BRAF* + *TERT* mutations with disease recurrence, advanced tumor stage, presence of metastases, and higher mortality risk (16, 47, 48, 49, 50, 51). *TERT* promoter mutations have also been linked to lower thyroid differentiation levels and subsequent higher refractoriness to standard radioiodine therapy in *BRAF*^{V600E}-driven PTC (11, 52, 53). Finally, *TERT* mutations also cooperate with *RAS*-driven DTCs toward tumor aggressiveness. Song *et al.* retrospectively assessed a group of 690 patients with FTC and showed

Table 1 Key studies using genomic characterization for differentiated thyroid cancer prognostication.

Reference	DTC type	DTC cohort, n	Gene(s) and mutations	Main findings
Ricarte-Filho <i>et al.</i> 2009 (66)	PTC, others	33	<i>AKT1</i> , <i>PIK3CA</i>	<i>AKT1</i> and <i>PIK3CA</i> mutations first reported in RAI refractory/metastatic PTCs, predominantly tall cell subtype
Melo <i>et al.</i> 2014 (45)	PTC + FTC	402	<i>TERT</i> promoter	<i>TERT</i> promoter mutations are associated with persistent and metastatic disease in PTC and FTC
Xing <i>et al.</i> 2014 (46)	PTC	507	<i>BRAF</i> ^{V600E} + <i>TERT</i> promoter	Co-occurrence of <i>BRAF</i> ^{V600E} and <i>TERT</i> ^{C>T} mutations associate with decreased recurrence-free survival of PTC patients
Song <i>et al.</i> 2016 (16)	PTC + FTC	551	<i>BRAF</i> ^{V600E} + <i>TERT</i> promoter; <i>RAS</i> + <i>TERT</i> promoter	Coexistence of <i>TERT</i> promoter mutations with either <i>BRAF</i> ^{V600E} or <i>RAS</i> alterations associate with increased disease-specific mortality
Pappa <i>et al.</i> 2021 (67)	PTC, <i>BRAF</i> ^{V600E} mutant	225	PI3K pathway (<i>PIK3CA</i> , <i>AKT1</i> , others)	Oncogenic mutations in PI3K pathway effectors are independent predictors of disease-specific mortality in <i>BRAF</i> ^{V600E} -driven PTCs
Nguyen <i>et al.</i> 2022 (76)	PTC, metastatic	307	<i>TERT</i> promoter, <i>TP53</i> , <i>CDKN2A</i> , others	Pancancer analysis of metastatic tumors shows higher frequencies of mutations in <i>TERT</i> , <i>TP53</i> and <i>CDKN2A</i> in metastatic PTC (compared to PTC profiled by the TCGA)
Boucai <i>et al.</i> 2023 (68)	PTC, FTC, struma ovarii	21	<i>RBM10</i> and <i>SWI/SNF</i> gene mutations; chromosome 1q gain	Alterations in <i>RBM10</i> , <i>SWI/SNF</i> members and chromosome 1q are enriched in nonresponders to RAI (compared to exceptional responders)
Bikas <i>et al.</i> 2023 (17)	PTC + FTC, <i>RAS</i> mutant	69	<i>TERT</i> promoter, <i>PTEN</i> , <i>SWI/SNF</i> genes, others	Presence of oncogenic mutation in specific genes predict disease-specific mortality in <i>RAS</i> -driven DTCs

DTC, differentiated thyroid cancer; FTC, follicular thyroid cancer; PTC, papillary thyroid cancer; RAI, radioactive iodine; TCGA, The Cancer Genome Atlas.

that combined *RAS* and *TERT* promoter mutations showed a higher recurrence risk than either of those mutations in isolation (54). Shen and colleagues, on their part, led a multicenter effort and demonstrated that PTC patients whose tumors harbored *RAS*+*TERT* mutations showed worse clinicopathological outcomes, notably higher risks of recurrence (55). Song *et al.* later studied a cohort of 551 DTCs, including 48 *RAS*-mutant tumors, and showed that, similarly to their *BRAF*^{V600E} + *TERT* counterparts, *RAS* + *TERT* mutations associated with decreased disease-free and disease-specific survival in DTC patients (compared to *RAS*-alone) (16).

Overall, the presence of a *TERT* promoter mutation is a bona fide marker of poor prognosis (for a detailed review and meta-analyses on this topic, see (56, 57, 58, 59)), and it is now gradually incorporated into the clinical guidelines for managing thyroid cancer patients (1).

RAS + EIF1AX mutations

Mutations in *EIF1AX*, a gene encoding a member of the translation initiation complex, were first reported in a small subset of the PTCs (2), and shortly after identified as much more frequent events in the PDTs and ATCs (28, 29). The pattern of mutations of *EIF1AX* in advanced diseased showed a remarkable association with the presence of *RAS* mutations, suggesting a cooperativity among those two genetic lesions that were later mechanistically dissected (60). *EIF1AX* alterations also occur in uveal melanomas (61), but a specific mutation

at a splice site at alanine 113 (A113splice) is exclusive of thyroid tumors.

Isolated *RAS* or *EIF1AX* mutations associate with follicular-patterned thyroid neoplasms, including benign follicular adenomas (FA), fvPTC, and FTC. The fact that the *RAS* + *EIF1AX* combination was enriched in PDTs and ATC prompted various groups to evaluate its role in the stratification of early disease. Karunamurthy and colleagues surveyed the prevalence of *EIF1AX* mutations across a series of thyroid specimens. They found that 4/4 carcinomas carrying an *EIF1AX* mutation did so at the A113splice position, and 3/4 had coexisting mutations in *NRAS* (62). More recently, three groups assessed the pattern of *EIF1AX* mutations in thyroid cancers. Gargano *et al.* reported, in a series of surgically removed nodules, a risk of malignancy of 100% for specimens with *EIF1AX* mutations (most of them at A113splice) when coexisting with additional mutations (overwhelmingly at *NRAS*, *HRAS*, or *KRAS*) (63). Karsoglu-French and colleagues evaluated 31 consecutive patients with *EIF1AX* mutations: the 18 specimens without coexisting mutations in *RAS* genes were enriched in benign disease (12/18 were FA, 3/18 hyperplastic nodules (HN) and 3/18 FTC), whereas the 13 *EIF1AX* + *RAS* samples showed primarily malignant phenotypes (6/13 PTC, 3/13 ATC, 2/13 NIFTP, 1/13 FA, and 1/13 HN) (64). Bandargal and colleagues, on their part, reported a multicenter series of 42 surgically resected nodules that harbored an *EIF1AX* mutation and showed that every single thyroid specimen with coexisting *EIF1AX* + *RAS* mutations was a carcinoma (65). Overall, there is growing evidence

suggesting that, although *RAS* or *EIF1AX* mutations can be found in isolation in a subset of FA, their co-occurrence likely pushes these lesions toward malignancy, typically prompting FTC, encapsulated or infiltrative fvPTC histotypes. Because of their low frequency in DTCs, so far no large studies have comprehensively evaluated whether *EIF1AX*-mutant tumors behave significantly differently than their counterparts with alternative genomic alterations.

Other mutations in DTC prognostication

The presence in DTCs of additional mutations in pathways that have been shown to promote cancer progression is rare but conveys clear prognostic significance. Ricarte-Filho *et al.* reported mutations in *AKT1* and *PIK3CA* (key effectors of the PI3K/AKT pathway) in BRAF^{V600E}-driven radioiodine-refractory metastatic/recurrent DTCs. PI3K/AKT pathway mutations were predominantly found in biopsies obtained from recurrent or metastatic sites and were enriched in PTCs with tall cell histotypes

(66). In a cohort of 225 BRAF^{V600E}-mutant PTCs, Pappa and colleagues showed that the presence of additional mutations in PI3K pathway effectors were independent predictors of disease-specific mortality (67). Song *et al.* confirmed the role of mutations in the PI3K pathway in increased mortality in a cohort of 50 BRAF^{V600E}-driven advanced PTCs and suggested that alterations in histone methyltransferase genes also had prognostic value (68). Bikas and colleagues, on their part, reported that additional mutations targeting several pathways predict disease-specific mortality in a cohort of 69 RAS-driven DTCs (17). In addition, two recent side-by-side publications from the same group demonstrated the power of comprehensive molecular testing in predicting tumor recurrence and refining the current methods for risk stratification of thyroid cancer patients (69, 70).

Regarding the molecular determinants of response to radioactive iodine (RAI) treatment, Boucai *et al.* compared a unique cohort of metastatic thyroid cancer patients with exceptional structural responses to RAI therapy vs matched nonresponders. Exceptional

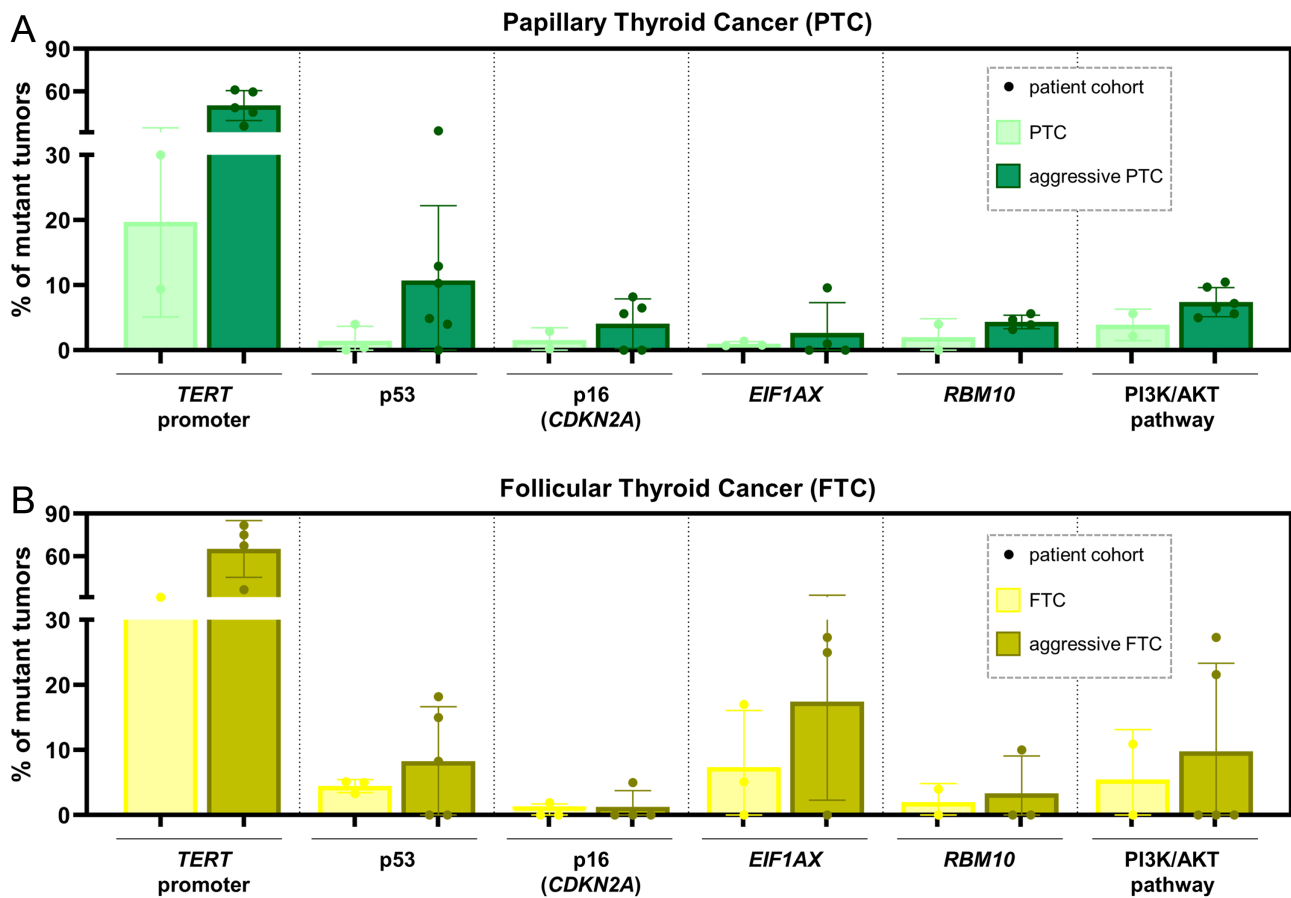


Figure 1

Comparison of the prevalence of mutations in key genes and pathways in differentiated thyroid cancer (DTC) patients vs DTC cohorts with aggressive/metastatic behavior in (A) papillary thyroid cancer (PTC) and (B) follicular thyroid cancer (FTC). Each dot represents an independent patient cohort. Data were obtained from studies employing next-generation sequencing technologies in nonoverlapping thyroid cancer cohorts between 2014 and 2022 (2, 30, 72, 73, 74, 75, 76, 77, 78, 79, 80). Only studies with at least ten patients were included to avoid small sample size bias.

responders were enriched in RAS-like alterations (e.g. mutations in RAS genes or in BRAF^{K601E}), whereas BRAF^{V600E} drove tumors from nonresponders. The latter also harbored *RBM10* and SWI/SNF gene mutations and chromosome 1q gains (71).

Besides the prognostic role of mutations in *loci* such as the *TERT* promoter, *EIF1AX*, or in effectors of the PI3K/AKT pathway in the context of BRAF- or RAS-mutant tumors, there is indirect evidence of other genetic alterations as biomarkers of DTC severity. These come from tumor sequencing studies in which rare subsets of PTC and FTC with aggressive/metastatic properties were evaluated (n (PTC)=18–379; n (FTC)=11–60 patients) (30, 72, 73, 74, 75) as well as from a recent pan-cancer study of metastatic disease which included 307 PTCs (76). In Fig. 1, we compared the prevalence of selected mutations, typically more frequent in PDTC/ATC, in aggressive DTC cohorts vs unselected DTC counterparts (typically more indolent) (2, 30, 72, 73, 74, 75, 76, 77, 78, 79, 80). The former comparison suggests that mutations in other *loci*, such as *TP53*, *CDKN2A* (p16), and *RBM10*, likely prime a subset of PTCs (Fig. 1A) and FTCs (Fig. 1B) toward more aggressive behavior.

Finally, apart from gene mutations, differences in the expression of messenger RNAs (mRNAs) and microRNAs (miRNAs) have been suggested as potential tools to stratify PTC patients. As detailed in the previous section, Boucai and colleagues stratified, based on thyroid differentiation scores, the 227 BRAF-mutant PTCs from the TCGA study into the BRAF-TDS-high vs the BRAF-TDS-low subgroups. The authors showed that the transcriptomes of the BRAF-TDS-high tumors were more akin to RAS-like PTCs, which tend to maintain the thyroid follicular architecture better. They also showed that, compared to BRAF-TDS-low, the BRAF-TDS-high group overexpressed genes related to cell polarity, such as *CDH16* and *PDHD1-L1*, as well as miRNAs targeting nodes in the TGF- β signaling pathway (11). Nieto *et al.* also analyzed the TCGA dataset, associated mRNA levels of *FN1*, *ITGA3*, and *MET* genes, as well as of miRNAs miR-486 and miR-1179 with recurrence of BRAF-like PTCs, and provided functional validation *in vitro* systems (81). The latter builds upon the abundant literature assessing the clinical and biological implications of specific miRNAs in thyroid cancer (82, 83, 84, 85, 86, 87), although so far, this knowledge is not applied in routine prognostication.

In summary, alterations in *BRAF*, *RAS*, and *RET* are initiating events in thyroid tumors and determine some of their features. However, the use of genomic information for prognostication purposes typically requires the evaluation of a larger set of mutations in other genes which have been shown to play a role in thyroid cancer progression. In this regard, mutations in the *TERT* promoter, *EIF1AX* (in RAS-mutant disease) and in effectors of the PI3K/AKT pathway are good candidates to become useful routine biomarkers in thyroid cancer prognostication. Advancing knowledge in prognostic implications of different mutations has helped clinicians

to individualize the care of patients with thyroid cancer. Intense follow-up visits and cross-sectional imaging are routinely recommended for patients whose tumors have specific combinations of oncogenic alterations, while less aggressive follow-up visits are usually recommended in patients with less aggressive mutations. Advancing knowledge on the genomic landscape of thyroid cancer has also led to the development of targeted systemic treatment in patients with advanced thyroid cancer in recent years.

Declaration of interest

Dr Sara Ahmadi has received research funding from Veracyte.

Funding

This work did not receive any specific grant from any funding agency in the public, commercial, or not-for-profit sector.

Author contribution statement

Dr Sara Ahmadi is a senior editor of *Endocrine Connections*. Dr Sara Ahmadi was not involved in the review or editorial process for this paper, on which she is listed as an author.

References

- Haugen BR, Alexander EK, Bible KC, Doherty GM, Mandel SJ, Nikiforov YE, Pacini F, Randolph GW, Sawka AM, Schlumberger M, *et al.* 2015 American Thyroid Association management guidelines for adult patients with thyroid nodules and differentiated thyroid cancer: the American Thyroid Association guidelines task force on thyroid nodules and differentiated thyroid cancer. *Thyroid* 2016 **26** 1–133. (<https://doi.org/10.1089/thy.2015.0020>)
- Cancer Genome Atlas Research Network. Integrated genomic characterization of papillary thyroid carcinoma. *Cell* 2014 **159** 676–690. (<https://doi.org/10.1016/j.cell.2014.09.050>)
- Li F, Chen G, Sheng C, Gusdon AM, Huang Y, Lv Z, Xu H, Xing M & Qu S. BRAFV600E mutation in papillary thyroid microcarcinoma: a meta-analysis. *Endocrine-Related Cancer* 2015 **22** 159–168. (<https://doi.org/10.1530/ERC-14-0531>)
- Tao Y, Wang F, Shen X, Zhu G, Liu R, Viola D, Elisei R, Puxeddu E, Fugazzola L, Colombo C, *et al.* BRAF V600E status sharply differentiates lymph node metastasis-associated mortality risk in papillary thyroid cancer. *Journal of Clinical Endocrinology and Metabolism* 2021 **106** 3228–3238. (<https://doi.org/10.1210/clinem/dgab286>)
- Walczyk A, Kowalska A, Kowalik A, Sygut J, Wypiorkiewicz E, Chodurska R, Pięciak L & Gózdź S. The BRAF(V600E) mutation in papillary thyroid microcarcinoma: does the mutation have an impact on clinical outcome? *Clinical Endocrinology* 2014 **80** 899–904. (<https://doi.org/10.1111/cen.12386>)
- Ito Y, Yoshida H, Maruo R, Morita S, Takano T, Hirokawa M, Yabuta T, Fukushima M, Inoue H, Tomoda C, *et al.* BRAF mutation in papillary thyroid carcinoma in a Japanese population: its lack of correlation with high-risk clinicopathological features and disease-free survival of patients. *Endocrine Journal* 2009 **56** 89–97. (<https://doi.org/10.1507/endocrj.k08e-208>)
- Choi SY, Park H, Kang MK, Lee DK, Lee KD, Lee HS, Kim SW, Lee EN & Hong JC. The relationship between the BRAF(V600E) mutation in

- papillary thyroid microcarcinoma and clinicopathologic factors. *World Journal of Surgical Oncology* 2013 **11** 291. (<https://doi.org/10.1186/1477-7819-11-291>)
- 8 Perera D, Ghossein R, Camacho N, Senbabaoglu Y, Seshan V, Li J, Bouvier N, Boucai L, Viale A, Socci ND, *et al.* Genomic and transcriptomic characterization of papillary microcarcinomas with lateral neck lymph node metastases. *Journal of Clinical Endocrinology and Metabolism* 2019 **104** 4889–4899. (<https://doi.org/10.1210/jc.2019-00431>)
 - 9 Gouveia C, Can NT, Bostrom A, Grenert JP, van Zante A & Orloff LA. Lack of association of BRAF mutation with negative prognostic indicators in papillary thyroid carcinoma: the University of California, San Francisco, experience. *JAMA Otolaryngology – Head and Neck Surgery* 2013 **139** 1164–1170. (<https://doi.org/10.1001/jamaoto.2013.4501>)
 - 10 Xing M, Alzahrani AS, Carson KA, Viola D, Elisei R, Bendlova B, Yip L, Mian C, Vianello F, Tuttle RM, *et al.* Association between BRAF V600E mutation and mortality in patients with papillary thyroid cancer. *JAMA* 2013 **309** 1493–1501. (<https://doi.org/10.1001/jama.2013.3190>)
 - 11 Boucai L, Seshan V, Williams M, Knauf JA, Saqcena M, Ghossein RA & Fagin JA. Characterization of subtypes of BRAF-mutant papillary thyroid cancer defined by their thyroid differentiation score. *Journal of Clinical Endocrinology and Metabolism* 2022 **107** 1030–1039. (<https://doi.org/10.1210/clinem/dgab851>)
 - 12 Rivera M, Tuttle RM, Patel S, Saha A, Shah JP & Ghossein RA. Encapsulated papillary thyroid carcinoma: a clinico-pathologic study of 106 cases with emphasis on its morphologic subtypes (histologic growth pattern). *Thyroid* 2009 **19** 119–127. (<https://doi.org/10.1089/thy.2008.0303>)
 - 13 Sabra MM, Dominguez JM, Grewal RK, Larson SM, Ghossein RA, Tuttle RM & Fagin JA. Clinical outcomes and molecular profile of differentiated thyroid cancers with radioiodine-avid distant metastases. *Journal of Clinical Endocrinology and Metabolism* 2013 **98** E829–E836. (<https://doi.org/10.1210/jc.2012-3933>)
 - 14 Nikiforov YE & Nikiforova MN. Molecular genetics and diagnosis of thyroid cancer. *Nature Reviews. Endocrinology* 2011 **7** 569–580. (<https://doi.org/10.1038/nrendo.2011.142>)
 - 15 Hudson TJ, Puzstaszeri MP, Hier MP, Forest VI, Yang JW & Payne RJ. Does the likelihood of malignancy in thyroid nodules with RAS mutations increase in direct proportion with the allele frequency percentage? *Journal of Otolaryngology – Head and Neck Surgery* 2023 **52** 12. (<https://doi.org/10.1186/s40463-022-00611-8>)
 - 16 Song YS, Lim JA, Choi H, Won JK, Moon JH, Cho SW, Lee KE, Park YJ, Yi KH, Park DJ, *et al.* Prognostic effects of tert promoter mutations are enhanced by coexistence with BRAF or RAS mutations and strengthen the risk prediction by the ATA or TNM staging system in differentiated thyroid cancer patients. *Cancer* 2016 **122** 1370–1379. (<https://doi.org/10.1002/cncr.29934>)
 - 17 Bikas A, Ahmadi S, Pappa T, Marqusee E, Wong K, Nehs MA, Cho NL, Haase J, Doherty GM, Sehgal K, *et al.* Additional oncogenic alterations in RAS-driven differentiated thyroid cancers associate with worse clinicopathologic outcomes. *Clinical Cancer Research* 2023 **29** 2678–2685. (<https://doi.org/10.1158/1078-0432.CCR-23-0278>)
 - 18 Lam AK, Montone KT, Nolan KA & Livolsi VA. Ret oncogene activation in papillary thyroid carcinoma: prevalence and implication on the histological parameters. *Human Pathology* 1998 **29** 565–568. ([https://doi.org/10.1016/s0046-8177\(98\)80004-x](https://doi.org/10.1016/s0046-8177(98)80004-x))
 - 19 Pisarchik AV, Ermak G, Demidchik EP, Mikhalevich LS, Kartel NA & Figue J. Low prevalence of the ret/PTC3r1 rearrangement in a series of papillary thyroid carcinomas presenting in Belarus ten years post-Chernobyl. *Thyroid* 1998 **8** 1003–1008. (<https://doi.org/10.1089/thy.1998.8.1003>)
 - 20 Thomas GA, Bunnell H, Cook HA, Williams ED, Nerovnya A, Cherstvoy ED, Tronko ND, Bogdanova TI, Chiappetta G, Viglietto G, *et al.* High prevalence of RET/PTC rearrangements in Ukrainian and Belarussian post-Chernobyl thyroid papillary carcinomas: a strong correlation between RET/PTC3 and the solid-follicular variant. *Journal of Clinical Endocrinology and Metabolism* 1999 **84** 4232–4238. (<https://doi.org/10.1210/jcem.84.11.6129>)
 - 21 Morton LM, Karyadi DM, Stewart C, Bogdanova TI, Dawson ET, Steinberg MK, Dai J, Hartley SW, Schonfeld SJ, Sampson JN, *et al.* Radiation-related genomic profile of papillary thyroid carcinoma after the Chernobyl accident. *Science* 2021 **372**. (<https://doi.org/10.1126/science.abg2538>)
 - 22 Ricarte-Filho JC, Li S, Garcia-Rendueles ME, Montero-Conde C, Voza F, Knauf JA, Heguy A, Viale A, Bogdanova T, Thomas GA, *et al.* Identification of kinase fusion oncogenes in post-Chernobyl radiation-induced thyroid cancers. *Journal of Clinical Investigation* 2013 **123** 4935–4944. (<https://doi.org/10.1172/JCI69766>)
 - 23 Rabes HM, Demidchik EP, Sidorow JD, Lengfelder E, Beimfohr C, Hoelzel D & Klugbauer S. Pattern of radiation-induced RET and NTRK1 rearrangements in 191 post-Chernobyl papillary thyroid carcinomas: biological, phenotypic, and clinical implications. *Clinical Cancer Research* 2000 **6** 1093–1103.
 - 24 Nikiforov YE, Rowland JM, Bove KE, Monforte-Munoz H & Fagin JA. Distinct pattern of ret oncogene rearrangements in morphological variants of radiation-induced and sporadic thyroid papillary carcinomas in children. *Cancer Research* 1997 **57** 1690–1694.
 - 25 Soares P, Fonseca E, Wynford-Thomas D & Sobrinho-Simoes M. Sporadic ret-rearranged papillary carcinoma of the thyroid: a subset of slow growing, less aggressive thyroid neoplasms? *Journal of Pathology* 1998 **185** 71–78. ([https://doi.org/10.1002/\(SICI\)1096-9896\(199805\)185:1<71::AID-PATH42>3.0.CO;2-S](https://doi.org/10.1002/(SICI)1096-9896(199805)185:1<71::AID-PATH42>3.0.CO;2-S))
 - 26 Tallini G, Santoro M, Helie M, Carlomagno F, Salvatore G, Chiappetta G, Carcangiu ML & Fusco A. RET/PTC oncogene activation defines a subset of papillary thyroid carcinomas lacking evidence of progression to poorly differentiated or undifferentiated tumor phenotypes. *Clinical Cancer Research* 1998 **4** 287–294.
 - 27 Franco AT, Ricarte-Filho JC, Isaza A, Jones Z, Jain N, Mostoufi-Moab S, Surrey L, Laetsch TW, Li MM, DeHart JC, *et al.* Fusion oncogenes are associated with increased metastatic capacity and persistent disease in pediatric thyroid cancers. *Journal of Clinical Oncology* 2022 **40** 1081–1090. (<https://doi.org/10.1200/JCO.21.01861>)
 - 28 Kunstman JW, Juhlin CC, Goh G, Brown TC, Stenman A, Healy JM, Rubinstein JC, Choi M, Kiss N, Nelson-Williams C, *et al.* Characterization of the mutational landscape of anaplastic thyroid cancer via whole-exome sequencing. *Human Molecular Genetics* 2015 **24** 2318–2329. (<https://doi.org/10.1093/hmg/ddu749>)
 - 29 Landa I, Ibrahimasic T, Boucai L, Sinha R, Knauf JA, Shah RH, Dogan S, Ricarte-Filho JC, Krishnamoorthy GP, Xu B, *et al.* Genomic and transcriptomic hallmarks of poorly differentiated and anaplastic thyroid cancers. *Journal of Clinical Investigation* 2016 **126** 1052–1066. (<https://doi.org/10.1172/JCI85271>)
 - 30 Pozdveyev N, Gay LM, Sokol ES, Hartmaier R, Deaver KE, Davis S, French JD, Borre PV, LaBarbera DV, Tan AC, *et al.* Genetic analysis of 779 advanced differentiated and anaplastic thyroid cancers. *Clinical Cancer Research* 2018 **24** 3059–3068. (<https://doi.org/10.1158/1078-0432.CCR-18-0373>)

- 31 Horn S, Figli A, Rachakonda PS, Fischer C, Sucker A, Gast A, Kadel S, Moll I, Nagore E, Hemminki K, *et al.* Tert promoter mutations in familial and sporadic melanoma. *Science* 2013 **339** 959–961. (<https://doi.org/10.1126/science.1230062>)
- 32 Huang FW, Hodis E, Xu MJ, Kryukov GV, Chin L & Garraway LA. Highly recurrent tert promoter mutations in human melanoma. *Science* 2013 **339** 957–959. (<https://doi.org/10.1126/science.1229259>)
- 33 Killela PJ, Reitman ZJ, Jiao Y, Bettgowda C, Agrawal N, Diaz LA, Friedman AH, Friedman H, Gallia GL, Giovannella BC, *et al.* Tert promoter mutations occur frequently in gliomas and a subset of tumors derived from cells with low rates of self-renewal. *PNAS* 2013 **110** 6021–6026. (<https://doi.org/10.1073/pnas.1303607110>)
- 34 Landa I, Ganly I, Chan TA, Mitsutake N, Matsuse M, Ibrahimspic T, Ghossein RA & Fagin JA. Frequent somatic tert promoter mutations in thyroid cancer: higher prevalence in advanced forms of the disease. *Journal of Clinical Endocrinology and Metabolism* 2013 **98** E1562–E1566. (<https://doi.org/10.1210/jc.2013-2383>)
- 35 Liu X, Bishop J, Shan Y, Pai S, Liu D, Murugan AK, Sun H, El-Naggar AK & Xing M. Highly prevalent tert promoter mutations in aggressive thyroid cancers. *Endocrine-Related Cancer* 2013 **20** 603–610. (<https://doi.org/10.1530/ERC-13-0210>)
- 36 Vinagre J, Almeida A, Populo H, Batista R, Lyra J, Pinto V, Coelho R, Celestino R, Prazeres H, Lima L, *et al.* Frequency of tert promoter mutations in human cancers. *Nature Communications* 2013 **4** 2185. (<https://doi.org/10.1038/ncomms3185>)
- 37 Fredriksson NJ, Ny L, Nilsson JA & Larsson E. Systematic analysis of noncoding somatic mutations and gene expression alterations across 14 tumor types. *Nature Genetics* 2014 **46** 1258–1263. (<https://doi.org/10.1038/ng.3141>)
- 38 Bullock M, Ren Y, O'Neill C, Gill A, Aniss A, Sywak M, Sidhu S, Delbridge L, Learoyd D, de Vathaire F, *et al.* Tert promoter mutations are a major indicator of recurrence and death due to papillary thyroid carcinomas. *Clinical Endocrinology* 2016 **85** 283–290. (<https://doi.org/10.1111/cen.12999>)
- 39 Liu R & Xing M. Diagnostic and prognostic tert promoter mutations in thyroid fine-needle aspiration biopsy. *Endocrine-Related Cancer* 2014 **21** 825–830. (<https://doi.org/10.1530/ERC-14-0359>)
- 40 Nikiforov YE, Carty SE, Chiosea SI, Coyne C, Duvvuri U, Ferris RL, Gooding WE, Hodak SP, LeBeau SO, Otori NP, *et al.* Highly accurate diagnosis of cancer in thyroid nodules with follicular neoplasm/suspicious for a follicular neoplasm cytology by ThyroSeq v2 next-generation sequencing assay. *Cancer* 2014 **120** 3627–3634. (<https://doi.org/10.1002/cncr.29038>)
- 41 Steward DL, Carty SE, Sippel RS, Yang SP, Sosa JA, Sapos JA, Figge JJ, Mandel S, Haugen BR, Burman KD, *et al.* Performance of a multigene genomic classifier in thyroid nodules with indeterminate cytology: a prospective blinded multicenter study. *JAMA Oncology* 2019 **5** 204–212. (<https://doi.org/10.1001/jamaoncol.2018.4616>)
- 42 Skaugen JM, Taneja C, Liu JB, Wald AI, Nikitski AV, Chiosea SI, Seethala RR, Otori NP, Karslioglu-French E, Carty SE, *et al.* Performance of a multigene genomic classifier in thyroid nodules with suspicious for malignancy cytology. *Thyroid* 2022 **32** 1500–1508. (<https://doi.org/10.1089/thy.2022.0282>)
- 43 Decaussin-Petrucci M, Descotes F, Depaepae L, Lapras V, Denier ML, Borson-Chazot F, Lifante JC & Lopez J. Molecular testing of BRAF, RAS and tert on thyroid FNAs with indeterminate cytology improves diagnostic accuracy. *Cytopathology* 2017 **28** 482–487. (<https://doi.org/10.1111/cyt.12493>)
- 44 Liu T, Wang N, Cao J, Sofiadis A, Dinets A, Zedenius J, Larsson C & Xu D. The age- and shorter telomere-dependent tert promoter mutation in follicular thyroid cell-derived carcinomas. *Oncogene* 2014 **33** 4978–4984. (<https://doi.org/10.1038/onc.2013.446>)
- 45 Melo M, da Rocha AG, Vinagre J, Batista R, Peixoto J, Tavares C, Celestino R, Almeida A, Salgado C, Eloy C, *et al.* Tert promoter mutations are a major indicator of poor outcome in differentiated thyroid carcinomas. *Journal of Clinical Endocrinology and Metabolism* 2014 **99** E754–E765. (<https://doi.org/10.1210/jc.2013-3734>)
- 46 Xing M, Liu R, Liu X, Murugan AK, Zhu G, Zeiger MA, Pai S & Bishop J. BRAF V600E and tert promoter mutations cooperatively identify the most aggressive papillary thyroid cancer with highest recurrence. *Journal of Clinical Oncology* 2014 **32** 2718–2726. (<https://doi.org/10.1200/JCO.2014.55.5094>)
- 47 Liu X, Qu S, Liu R, Sheng C, Shi X, Zhu G, Murugan AK, Guan H, Yu H, Wang Y, *et al.* Tert promoter mutations and their association with BRAF V600E mutation and aggressive clinicopathological characteristics of thyroid cancer. *Journal of Clinical Endocrinology and Metabolism* 2014 **99** E1130–E1136. (<https://doi.org/10.1210/jc.2013-4048>)
- 48 Gandolfi G, Ragazzi M, Frasoldati A, Piana S, Ciarrocchi A & Sancisi V. Tert promoter mutations are associated with distant metastases in papillary thyroid carcinoma. *European Journal of Endocrinology* 2015 **172** 403–413. (<https://doi.org/10.1530/EJE-14-0837>)
- 49 Liu R, Bishop J, Zhu G, Zhang T, Ladenson PW & Xing M. Mortality risk stratification by combining BRAF V600E and tert promoter mutations in papillary thyroid cancer: genetic duet of BRAF and tert promoter mutations in thyroid cancer mortality. *JAMA Oncology* 2017 **3** 202–208. (<https://doi.org/10.1001/jamaoncol.2016.3288>)
- 50 Lee SE, Hwang TS, Choi YL, Han HS, Kim WS, Jang MH, Kim SK & Yang JH. Prognostic significance of tert promoter mutations in papillary thyroid carcinomas in a BRAF(V600E) mutation-prevalent population. *Thyroid* 2016 **26** 901–910. (<https://doi.org/10.1089/thy.2015.0488>)
- 51 Povoaa AA, Teixeira E, Bella-Cueto MR, Batista R, Pestana A, Melo M, Alves T, Pinto M, Sobrinho-Simões M, Maciel J, *et al.* Genetic determinants for prediction of outcome of patients with papillary thyroid carcinoma. *Cancers* 2021 **13**. (<https://doi.org/10.3390/cancers13092048>)
- 52 Liu J, Liu R, Shen X, Zhu G, Li B & Xing M. The genetic duet of BRAF V600E and tert promoter mutations robustly predicts loss of radioiodine avidity in recurrent papillary thyroid cancer. *Journal of Nuclear Medicine* 2020 **61** 177–182. (<https://doi.org/10.2967/jnumed.119.227652>)
- 53 Soe MH, Chiang JM, Flavell RR, Khanafshar E, Mendoza L, Kang H & Liu C. Non-iodine-avid disease is highly prevalent in distant metastatic differentiated thyroid cancer with papillary histology. *Journal of Clinical Endocrinology and Metabolism* 2022 **107** e3206–e3216. (<https://doi.org/10.1210/clinem/dgac305>)
- 54 Song YS, Lim JA, Min HS, Kim MJ, Choi HS, Cho SW, Moon JH, Yi KH, Park DJ, Cho BY, *et al.* Changes in the clinicopathological characteristics and genetic alterations of follicular thyroid cancer. *European Journal of Endocrinology* 2017 **177** 465–473. (<https://doi.org/10.1530/EJE-17-0456>)
- 55 Shen X, Liu R & Xing M. A six-genotype genetic prognostic model for papillary thyroid cancer. *Endocrine-Related Cancer* 2017 **24** 41–52. (<https://doi.org/10.1530/ERC-16-0402>)
- 56 Alzahrani AS, Alsaadi R, Murugan AK & Sadiq BB. Tert promoter mutations in thyroid cancer. *Hormones and Cancer* 2016 **7** 165–177. (<https://doi.org/10.1007/s12672-016-0256-3>)
- 57 Moon S, Song YS, Kim YA, Lim JA, Cho SW, Moon JH, Hahn S, Park DJ & Park YJ. Effects of coexistent BRAF(V600E) and tert promoter mutations on poor clinical outcomes in papillary thyroid cancer: a meta-analysis. *Thyroid* 2017 **27** 651–660. (<https://doi.org/10.1089/thy.2016.0350>)

- 58 Chen B, Shi Y, Xu Y & Zhang J. The predictive value of coexisting BRAFV600E and tert promoter mutations on poor outcomes and high tumour aggressiveness in papillary thyroid carcinoma: a systematic review and meta-analysis. *Clinical Endocrinology* 2021 **94** 731–742. (<https://doi.org/10.1111/cen.14316>)
- 59 Zhao L, Wang L, Jia X, Hu X, Pang P, Zhao S, Wang Y, Wang J, Zhang Y & Lyu Z. The coexistence of genetic mutations in thyroid carcinoma predicts histopathological factors associated with a poor prognosis: a systematic review and network meta-analysis. *Frontiers in Oncology* 2020 **10** 540238. (<https://doi.org/10.3389/fonc.2020.540238>)
- 60 Krishnamoorthy GP, Davidson NR, Leach SD, Zhao Z, Lowe SW, Lee G, Landa I, Nagarajah J, Saqcena M, Singh K, et al. EIF1AX and RAS mutations cooperate to drive thyroid tumorigenesis through ATF4 and c-MYC. *Cancer Discovery* 2019 **9** 264–281. (<https://doi.org/10.1158/2159-8290.CD-18-0606>)
- 61 Martin M, Masshofer L, Temming P, Rahmann S, Metz C, Bornfeld N, van de Nes J, Klein-Hitpass L, Hinnebusch AG, Horsthemke B, et al. Exome sequencing identifies recurrent somatic mutations in EIF1AX and SF3B1 in uveal melanoma with disomy 3. *Nature Genetics* 2013 **45** 933–936. (<https://doi.org/10.1038/ng.2674>)
- 62 Karunamurthy A, Panebianco F, Hsiao SJ, Vorhauer J, Nikiforova MN, Chiosea S & Nikiforov YE. Prevalence and phenotypic correlations of EIF1AX mutations in thyroid nodules. *Endocrine-Related Cancer* 2016 **23** 295–301. (<https://doi.org/10.1530/ERC-16-0043>)
- 63 Gargano SM, Badjatia N, Nikolaus Y, Peiper SC & Wang ZX. Characterization and clinical significance of EIF1AX mutations and co-mutations in cytologically indeterminate thyroid nodules: a 5-year retrospective analysis. *Acta Medica Academica* 2021 **50** 4–12. (<https://doi.org/10.5644/ama2006-124.322>)
- 64 Karslioglu French E, Nikitski AV, Yip L, Nikiforova MN, Nikiforov YE & Carty SE. Clinicopathological features and outcomes of thyroid nodules with EIF1AX mutations. *Endocrine-Related Cancer* 2022 **29** 467–473. (<https://doi.org/10.1530/ERC-22-0041>)
- 65 Bandargal S, Chen T, Pusztaszeri MP, Forest VI, da Silva SD & Payne RJ. Prognostic indicators of EIF1AX-mutated thyroid tumor malignancy and cancer aggressiveness. *Cancers* 2022 **14**. (<https://doi.org/10.3390/cancers14246097>)
- 66 Ricarte-Filho JC, Ryder M, Chitale DA, Rivera M, Heguy A, Ladanyi M, Janakiraman M, Solit D, Knauf JA, Tuttle RM, et al. Mutational profile of advanced primary and metastatic radioactive iodine-refractory thyroid cancers reveals distinct pathogenetic roles for BRAF, PIK3CA, and AKT1. *Cancer Research* 2009 **69** 4885–4893. (<https://doi.org/10.1158/0008-5472.CAN-09-0727>)
- 67 Pappa T, Ahmadi S, Marqusee E, Johnson HL, Nehs MA, Cho NL, Barletta JA, Lorch JH, Doherty GM, Lindeman NI, et al. Oncogenic mutations in PI3K/AKT/mTOR pathway effectors associate with worse prognosis in BRAF(V600E)-Driven papillary thyroid cancer patients. *Clinical Cancer Research* 2021 **27** 4256–4264. (<https://doi.org/10.1158/1078-0432.CCR-21-0874>)
- 68 Song E, Jin M, Jang A, Jeon MJ, Song DE, Yoo HJ, Kim WB, Shong YK & Kim WG. Mutation in genes encoding key functional groups additively increase mortality in patients with BRAF(V600E)-mutant advanced papillary thyroid carcinoma. *Cancers* 2021 **13**. (<https://doi.org/10.3390/cancers13225846>)
- 69 Liu JB, Ramonell KM, Carty SE, McCoy KL, Schaitkin BM, Karslioglu-French E, Morariu EM, Ohori NP, Seethala RR, Chiosea SI, et al. Association of comprehensive thyroid cancer molecular profiling with tumor phenotype and cancer-specific outcomes. *Surgery* 2023 **173** 252–259. (<https://doi.org/10.1016/j.surg.2022.05.048>)
- 70 Liu JB, Baugh KA, Ramonell KM, McCoy KL, Karslioglu-French E, Morariu EM, Ohori NP, Nikiforova MN, Nikiforov YE, Carty SE, et al. Molecular testing predicts incomplete response to initial therapy in differentiated thyroid carcinoma without lateral neck or distant metastasis at presentation: retrospective cohort study. *Thyroid* 2023 **33** 705–714. (<https://doi.org/10.1089/thy.2023.0060>)
- 71 Boucai L, Saqcena M, Kuo F, Grewal RK, Socci N, Knauf JA, Krishnamoorthy GP, Ryder M, Ho AL, Ghossein RA, et al. Genomic and transcriptomic characteristics of metastatic thyroid cancers with exceptional responses to radioactive iodine therapy. *Clinical Cancer Research* 2023 **29** 1620–1630. (<https://doi.org/10.1158/1078-0432.CCR-22-2882>)
- 72 Yoo SK, Song YS, Lee EK, Hwang J, Kim HH, Jung G, Kim YA, Kim SJ, Cho SW, Won JK, et al. Integrative analysis of genomic and transcriptomic characteristics associated with progression of aggressive thyroid cancer. *Nature Communications* 2019 **10** 2764. (<https://doi.org/10.1038/s41467-019-10680-5>)
- 73 Song E, Song DE, Ahn J, Kim TY, Kim WB, Shong YK, Jeon MJ & Kim WG. Genetic profile of advanced thyroid cancers in relation to distant metastasis. *Endocrine-Related Cancer* 2020 **27** 285–293. (<https://doi.org/10.1530/ERC-19-0452>)
- 74 Ibrahimspasic T, Xu B, Landa I, Dogan S, Middha S, Seshan V, Deraje S, Carlson DL, Migliacci J, Knauf JA, et al. Genomic alterations in fatal forms of non-anaplastic thyroid cancer: identification of MED12 and RBM10 as novel thyroid cancer genes associated with tumor virulence. *Clinical Cancer Research* 2017 **23** 5970–5980. (<https://doi.org/10.1158/1078-0432.CCR-17-1183>)
- 75 Paulsson JO, Rafati N, DiLorenzo S, Chen Y, Haglund F, Zedenius J & Juhlin CC. Whole-genome sequencing of follicular thyroid carcinomas reveal recurrent mutations in microRNA processing subunit DGCR8. *Journal of Clinical Endocrinology and Metabolism* 2021 **106** 3265–3282. (<https://doi.org/10.1210/clinem/dgab471>)
- 76 Nguyen B, Fong C, Luthra A, Smith SA, DiNatale RG, Nandakumar S, Walch H, Chatila WK, Madupuri R, Kundra R, et al. Genomic characterization of metastatic patterns from prospective clinical sequencing of 25,000 patients. *Cell* 2022 **185** 563–575.e11. (<https://doi.org/10.1016/j.cell.2022.01.003>)
- 77 AACR Project GENIE Consortium. AACR project GENIE: powering precision medicine through an international Consortium. *Cancer Discovery* 2017 **7** 818–831. (<https://doi.org/10.1158/2159-8290.CD-17-0151>)
- 78 Yoo SK, Lee S, Kim SJ, Jee HG, Kim BA, Cho H, Song YS, Cho SW, Won JK, Shin JY, et al. Comprehensive analysis of the transcriptional and mutational landscape of follicular and papillary thyroid cancers. *PLoS Genetics* 2016 **12** e1006239. (<https://doi.org/10.1371/journal.pgen.1006239>)
- 79 Nicolson NG, Murtha TD, Dong W, Paulsson JO, Choi J, Barbieri AL, Brown TC, Kunstman JW, Larsson C, Prasad ML, et al. Comprehensive genetic analysis of follicular thyroid carcinoma predicts prognosis independent of histology. *Journal of Clinical Endocrinology and Metabolism* 2018 **103** 2640–2650. (<https://doi.org/10.1210/jc.2018-00277>)
- 80 Chen H, Luthra R, Routbort MJ, Patel KP, Cabanillas ME, Broaddus RR & Williams MD. Molecular profile of advanced thyroid carcinomas by next-generation sequencing: characterizing tumors beyond diagnosis for targeted therapy. *Molecular Cancer Therapeutics* 2018 **17** 1575–1584. (<https://doi.org/10.1158/1535-7163.MCT-17-0871>)
- 81 Nieto HR, Thornton CEM, Brookes K, Nobre de Menezes A, Fletcher A, Alshahrani M, Kocbiyik M, Sharma N, Boelaert K, Cazier JB, et al. Recurrence of papillary thyroid cancer: a systematic appraisal of risk factors. *Journal of Clinical Endocrinology and*

- Metabolism* 2022 **107** 1392–1406. (<https://doi.org/10.1210/clinem/dgab836>)
- 82 Chou CK, Chen RF, Chou FF, Chang HW, Chen YJ, Lee YF, Yang KD, Cheng JT, Huang CC & Liu RT. miR-146b is highly expressed in adult papillary thyroid carcinomas with high risk features including extrathyroidal invasion and the BRAF(V600E) mutation. *Thyroid* 2010 **20** 489–494. (<https://doi.org/10.1089/thy.2009.0027>)
- 83 Riesco-Eizaguirre G, Wert-Lamas L, Perales-Paton J, Sastre-Perona A, Fernandez LP & Santisteban P. The miR-146b-3p/PAX8/NIS regulatory circuit modulates the differentiation phenotype and function of thyroid cells during carcinogenesis. *Cancer Research* 2015 **75** 4119–4130. (<https://doi.org/10.1158/0008-5472.CAN-14-3547>)
- 84 Han PA, Kim HS, Cho S, Fazeli R, Najafian A, Khawaja H, McAlexander M, Dy B, Sorensen M, Aronova A, *et al.* Association of BRAF V600E mutation and microRNA expression with central lymph node metastases in papillary thyroid cancer: a prospective study from four endocrine surgery centers. *Thyroid* 2016 **26** 532–542. (<https://doi.org/10.1089/thy.2015.0378>)
- 85 Mancikova V, Castelblanco E, Pineiro-Yanez E, Perales-Paton J, de Cubas AA, Inglada-Perez L, Matias-Guiu X, Capel I, Bella M, Lerma E, *et al.* MicroRNA deep-sequencing reveals master regulators of follicular and papillary thyroid tumors. *Modern Pathology* 2015 **28** 748–757. (<https://doi.org/10.1038/modpathol.2015.44>)
- 86 Dettmer M, Perren A, Moch H, Komminoth P, Nikiforov YE & Nikiforova MN. Comprehensive microRNA expression profiling identifies novel markers in follicular variant of papillary thyroid carcinoma. *Thyroid* 2013 **23** 1383–1389. (<https://doi.org/10.1089/thy.2012.0632>)
- 87 Ramirez-Moya J, Wert-Lamas L & Santisteban P. MicroRNA-146b promotes PI3K/AKT pathway hyperactivation and thyroid cancer progression by targeting PTEN. *Oncogene* 2018 **37** 3369–3383. (<https://doi.org/10.1038/s41388-017-0088-9>)