Interleukin-18 serum level is elevated in type 2 diabetes and latent autoimmune diabetes

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Abstract

Background: Interleukin-18 (IL-18) is an inflammatory cytokine found to be elevated in obesity, metabolic syndrome and type 2 diabetes (T2D) as a part of the chronic low-grade inflammatory process in these states. The aim of the study was to evaluate the interleukin level in patients with latent autoimmune diabetes of the adults (LADA) in comparison to that in T2D subjects.

Materials and methods: IL-18 was analyzed through enzyme-linked immunosorbent assay in 76 participants with T2D and 24 with LADA and 14 control subjects. Evaluation was also carried out in body mass index (BMI)- and glycemic control-matched diabetic patients.

Results: The serum concentration of IL-18 was higher in patients with T2D (389.04 ± 203.44 pg/mL) and LADA (327.04 ± 144.48 pg/mL) than that in control subjects (219.88 ± 91.03 pg/mL), P < 0.05. However, it was not significantly different between both diabetic groups (P = 0.255) despite higher IL-6 (4.78 ± 5.84 vs 1.79 ± 0.96 pg/mL, P < 0.001) and hs-CRP (2.60 ± 1.70 vs 1.29 ± 1.20 mg/L, P = 0.002) level in T2D patients. The results were persistent in BMI-matched subjects with diabetes (IL-18 = 403.48 ± 226.32 vs 329.30 ± 146.30 pg/mL, respectively for T2D and LADA, P = 0.391). The correlations in T2D group concerning HDL cholesterol (r = −0.377, P = 0.001), postprandial glucose (r = 0.244, P = 0.043), IL-6 (r = 0.398, P < 0.001) and hs-CRP (r = 0.427, P = 0.001) were not confirmed in LADA and control subjects. Conclusion: The IL-18 serum level was higher in T2D and LADA than that in control subjects, but did not differ between both diabetic groups, even when they were BMI matched. Correlations with lipid, glycemic and inflammatory parameters were present in T2D only.

Introduction

Interleukin-18 (IL-18), discovered in the late 20th century as an interferon-gamma (IFNγ)-inducing factor (1), is a cytokine that belongs to the IL-1 superfamily (2). It is produced by monocyte/macrophages, endothelial cells, smooth muscle cells, etc. and stimulates the expression of adhesion molecules, chemokine receptors, granulocyte macrophage colony-stimulating factor (GM-CSF), IFNγ, tumor necrosis factor α (TNFα) and interleukin 1β (IL-1β)
and activates type 1 or type 2 T helper immune response (3, 4). Apart from its place in immune defense against infective pathogens, IL-18 participates in the pathogenesis of atopic, autoimmune and chronic inflammatory diseases (3, 4).

Obesity and type 2 diabetes (T2D) have been associated with a chronic low-grade inflammation (5) that has been implicated as a crucial factor for insulin resistance (6) and also beta-cell dysfunction (7, 8). Patients diagnosed with latent autoimmune diabetes of the adults (LADA) (9), considered by many an indistinguishable part of type 1 diabetes (T1D) (10, 11, 60), usually present with a better metabolic profile and inflammatory markers than T2D, even though optimal glycemic control is usually harder to achieve (12).

Like other pro-inflammatory cytokines (such as IL-1β, TNFα, interleukin 6 (IL-6) (13, 14, 15, 16) and highsensitivity C-reactive protein (hs-CRP) (17)), elevated IL-18 level is a characteristic of obesity and the metabolic syndrome (18, 19, 20), and is reduced by weight loss (19, 21). The presence of the metabolic syndrome is associated with even higher IL-18 concentration than obesity alone, as has been shown for IL-6, TNFα and CRP (22). It is now well known that adipose tissue is a major contributor to inflammatory cytokine release in these states (23). As with other adipokines (24, 25), adipocytes can produce IL-18 (26, 27), but they do not seem to be the main source of the interleukin – rather it is being predominantly released by infiltrating macrophages (28).

Elevated IL-18 plasma level has been observed in vivo in response to acute hyperglycemia in healthy volunteers and subjects with impaired glucose tolerance (29). Patients with newly diagnosed T2D have been shown to have a significantly higher IL-18 level in comparison to non-diabetic subjects (30, 31). This elevation correlates with various factors used in the assessment of the metabolic risk – BMI, waist circumference, HDL cholesterol, triglycerides, blood pressure control, basal insulin, fasting plasma glucose (31, 32, 33) and insulin resistance index (34). Changes in IL-18 seem to be predictive of the risk of prediabetes (35) and T2D (35, 36, 37) in a way that is independent of other markers that reflect the ongoing chronic subclinical inflammation (37). They also contribute to determining the cardiovascular risk in the presence of a metabolic syndrome (38).

The aim of the current study was to compare the serum level of IL-18 between patients with T2D and subjects who meet the criteria for LADA (39) and to estimate its association with weight, glycemic and lipid control, as well as that of the inflammatory markers IL-6 and hs-CRP within both groups.

Subjects and methods

One hundred patients with diabetes were enrolled in this study – 76 patients with T2D and 24 with LADA and 14 control participants. The protocol of the study was in accordance with the declaration of Helsinki (40) and was approved by the ethics committee of Medical University-Sofia. All participants have signed an informed consent. Patients with LADA were enrolled if medical history included all of the following criteria: (1) onset at the age of 35 years or over; (2) presence of positive diabetes-associated autoantibodies, such as islet-cell cytoplasm autoantibodies (ICA), glutamic acid decarboxylase autoantibodies (GAD65A) (41), insulinoma-associated-2 antibodies – tyrosine phosphatase-associated (IA-2A) (42), zinc transporter 8 autoantibodies (ZnT8A) (43); (3) no need for insulin treatment for at least 6 months after diagnosis (39). All participants were assayed for GAD65A within the study as well (Supplementary Table 1, see section on supplementary data given at the end of this article).

The control groups consisted of healthy volunteers with a BMI <30 kg/m². Presence of the metabolic syndrome was assessed through the International Diabetes Federation criteria (44).

Blood samples were obtained after an overnight fasting state. Lipid profile, fasting plasma glucose (FPG) and 2-h post standard lunch meal postprandial plasma glucose (PPG), HbA1c were assessed by standard techniques in the Central Laboratory of the University hospital, which is the referent hospital for Bulgaria. IL-6 was analyzed by electro-chemiluminescence immunoassay on Roche Elecsys 2010 (Roche Diagnostics GmbH) and hs-CRP by particle enhanced immunoturbidimetric assay on Cobas Integra 400 Plus (Roche Diagnostics GmbH) with a lower detection limit of 1.5 pg/mL and 0.15 mg/L, respectively.

IL-18 was analyzed by enzyme-linked immunosorbent assay (ELISA) (Medical & Biological Laboratories Co., Ltd, Nagoya, Japan) with a detection limit of 12.5 pg/mL. GAD65A were assayed by ELISA (Euroimmune Medizinische Labordiagnostika AG, Lübeck, Germany) with a diagnostic cut-off at 10 IU/mL and a sensitivity and specificity, evaluated in the 2005 Diabetes Autoantibody Standardization Program workshop, of 92% and 98%, respectively.
Statistical analysis

Statistical analysis was performed with SPSS 21 statistics package. Comparison of data between independent samples was analyzed through the Mann–Whitney U test. Spearman’s rank correlation coefficient was computed for correlations. Fisher’s exact test was used for assessing differences in frequencies between groups. P values less than 0.05 were considered significant.

Results

Subjects’ characteristics are detailed in Table 1. Patients from both diabetic groups had a significantly higher serum level of IL-18 than the control participants. Taking into consideration the significant difference in gender distribution between T2D subjects and control participants, re-evaluation of the serum level only in women from these groups confirmed a significantly higher serum concentration of the IL-18 in T2D (n=43) than in control (n=13) participants (400 ± 197 vs 222 ± 94 pg/mL, P<0.001).

Despite the subjects with T2D and LADA having significantly different anthropometric parameters and glycemic control, and the T2D group having a higher IL-6 and hs-CRP level, the IL-18 level was not significantly different between both diabetic groups. Analysis between BMI-matched diabetic patients, who still differed in some parameters used in the assessment of metabolic risk (44) but had similar glycemic control, showed that the IL-6 and hs-CRP, although not significantly for the latter, remained higher in T2D group. The difference in the level of IL-18 was still not significant between both diabetic groups (Table 1).

Comparison in IL-18 level between BMI-matched T2D (n=8) and control (n=13) participants (BMI = 22.51 ± 1.08 vs 21.47 ± 1.73 kg/m², P=0.214; WC = 82 ± 8 vs 78 ± 8 cm, P=0.50) demonstrated a persistent higher level of the cytokine in the diabetic than that in control subjects (IL-18 = 430 ± 224 vs 218 ± 94 pg/mL, P<0.001).

This was also observed (IL-18 = 326 ± 135 vs 220 ± 91 pg/mL, P=0.027) between the LADA cohort (n=14) and control participants (n=14) when anthropometric parameters were not significantly different (BMI = 22.63 ± 2.40 vs 21.72 ± 1.91 kg/m², P=0.214; WC = 83 ± 8 vs 77 ± 8 cm, P=0.054).

The correlation analysis showed significant findings in the T2D group concerning relationship between the IL-18 level and glycemic and lipid control, but not

Table 1: Characteristics of participants.

<table>
<thead>
<tr>
<th>Characteristics of participants</th>
<th>T2D (n=28)</th>
<th>LADA (n=14)</th>
<th>Controls (n=13)</th>
<th>P</th>
<th>P*</th>
<th>P**</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gender (m/f)</td>
<td>16/12</td>
<td>8/6</td>
<td>5/8</td>
<td>&lt;0.001</td>
<td>0.067</td>
<td>0.003</td>
</tr>
<tr>
<td>Age (years)</td>
<td>54 ± 8</td>
<td>54 ± 6</td>
<td>51 ± 10</td>
<td>0.391</td>
<td>0.677</td>
<td>0.677</td>
</tr>
<tr>
<td>Duration (years)</td>
<td>4 ± 3</td>
<td>3 ± 2</td>
<td>3 ± 2</td>
<td>0.147</td>
<td>0.600</td>
<td>0.983</td>
</tr>
<tr>
<td>Height (cm)</td>
<td>170 ± 10</td>
<td>171 ± 8</td>
<td>173 ± 9</td>
<td>0.391</td>
<td>0.677</td>
<td>0.677</td>
</tr>
<tr>
<td>Weight (kg)</td>
<td>75 ± 12</td>
<td>68 ± 10</td>
<td>57 ± 10</td>
<td>0.041</td>
<td>0.214</td>
<td>0.147</td>
</tr>
<tr>
<td>BMI (kg/m²)</td>
<td>34.7 ± 5</td>
<td>27.4 ± 6</td>
<td>21.6 ± 1.8</td>
<td>&lt;0.001</td>
<td>0.014</td>
<td>0.007</td>
</tr>
<tr>
<td>WC (cm)</td>
<td>97 ± 10</td>
<td>82 ± 8</td>
<td>73 ± 8</td>
<td>&lt;0.001</td>
<td>0.014</td>
<td>0.007</td>
</tr>
<tr>
<td>SBP (mmHg)</td>
<td>136 ± 18</td>
<td>121 ± 12</td>
<td>114 ± 12</td>
<td>&lt;0.001</td>
<td>0.014</td>
<td>0.007</td>
</tr>
<tr>
<td>DBP (mmHg)</td>
<td>84 ± 11</td>
<td>73 ± 9</td>
<td>68 ± 9</td>
<td>&lt;0.001</td>
<td>0.014</td>
<td>0.007</td>
</tr>
<tr>
<td>FPG (mmol/L)</td>
<td>8.2 ± 3</td>
<td>7.2 ± 2</td>
<td>5.0 ± 0.3</td>
<td>&lt;0.001</td>
<td>0.014</td>
<td>0.007</td>
</tr>
<tr>
<td>PPG (mmol/L)</td>
<td>8.4 ± 3</td>
<td>10.4 ± 4</td>
<td>9.4 ± 3</td>
<td>0.041</td>
<td>0.214</td>
<td>0.147</td>
</tr>
<tr>
<td>HbA1c (%)</td>
<td>7.9 ± 2</td>
<td>9.2 ± 1</td>
<td>8.7 ± 2</td>
<td>0.001</td>
<td>0.014</td>
<td>0.007</td>
</tr>
<tr>
<td>TC (mmol/L)</td>
<td>5.2 ± 1</td>
<td>5.3 ± 1</td>
<td>5.0 ± 0.8</td>
<td>0.214</td>
<td>0.067</td>
<td>0.067</td>
</tr>
<tr>
<td>LDL-C (mmol/L)</td>
<td>3.0 ± 1</td>
<td>2.7 ± 1</td>
<td>2.5 ± 0.9</td>
<td>0.214</td>
<td>0.067</td>
<td>0.067</td>
</tr>
<tr>
<td>HDL-C (mmol/L)</td>
<td>1.3 ± 0.4</td>
<td>1.7 ± 0.5</td>
<td>1.8 ± 0.4</td>
<td>0.214</td>
<td>0.067</td>
<td>0.067</td>
</tr>
<tr>
<td>TG (mmol/L)</td>
<td>1.8 ± 1</td>
<td>1.4 ± 0.9</td>
<td>1.2 ± 0.8</td>
<td>0.214</td>
<td>0.067</td>
<td>0.067</td>
</tr>
<tr>
<td>hs-CRP (mg/L)</td>
<td>2.4 ± 1</td>
<td>1.3 ± 0.6</td>
<td>1.9 ± 0.6</td>
<td>0.214</td>
<td>0.067</td>
<td>0.067</td>
</tr>
</tbody>
</table>

*Compared with T2D; **compared with LADA; #characteristics within groups with BMI-matched participants.

BMI, body mass index; DBP, diastolic blood pressure; FPG, fasting plasma glucose; HDL-C, high density lipoprotein cholesterol; HbA1c, hemoglobin A1c; IL-6, interleukin-6; IL-18, interleukin-18; LADA, latent autoimmune diabetes of the adults; LDL-C, low density lipoprotein cholesterol; MS, metabolic syndrome; PPG, postprandial plasma glucose; SBP, systolic blood pressure; T2D, type 2 diabetes; TC, total cholesterol; TG, triglycerides; WC, waist circumference.
Obesity, as assessed by both BMI and waist circumference. However, IL-18 correlated positively with both IL-6 and hs-CRP serum concentrations only in T2D group. Results concerning IL-6, hs-CRP and HDL cholesterol were similar even when analysis included subjects with lower BMI. None of these results was observed in the LADA and control groups (Table 2).

**Discussion**

Chronic low-grade inflammation is part of the pathogenesis of T2D (5), while LADA seems to be characterized with lower levels of inflammatory cytokines like IL-6 and TNF-α (45). Serine phosphorylation of insulin receptor substrate (IRS), inhibition of the tyrosine kinase activity of the insulin receptor, ubiquitination and degradation of both IRS1 and IRS2, decreased transcription of IRS1 mRNA are some of the mechanisms involved in the reduction of insulin sensitivity in hepatocytes, adipocytes and muscles by the increased level of inflammatory cytokines (46). Whether IL-18 contributes to insulin resistance (47) is still not well understood although it has been associated with signal transducer and activator of transcription (STAT) signaling (48). Although insulin resistance cannot be excluded in some cases of autoimmune diabetes (49, 50), the metabolic syndrome and associated chronic inflammation are less often observed than in T2D (12, 45).

Unchanged serum concentration of IL-18 in LADA in comparison to T2D could be attributed to the stimulatory effect of hyperglycemia (51), and also to the involvement of the cytokine in autoimmune diseases, including diabetes mellitus (52). Increased level of IL-18 has been observed in some autoimmune diseases as multiple sclerosis, myasthenia gravis, systemic lupus erythematosus, Sjögren syndrome and rheumatoid arthritis, in both experimental models and clinical practice (52). It has been found to be elevated in T1D as well and its higher expression in the islets could be involved in the autoimmune response against beta-cells (51, 53). We have not been able to find data about the serum concentration of IL-18 in LADA in the literature, but being considered a part of the spectrum of T1D (10, 11, 60), we cannot exclude that its level in circulation is influenced by autoimmune processes in the islets.

It has been previously shown that IL-6 and TNF-α levels differ significantly in subjects with T2D and patients with late-onset diabetes and positive anti-GAD65 autoantibodies (45) and we have also confirmed different inflammatory parameters in both states (54). Despite the expected higher serum concentrations of IL-6 and hs-CRP in T2D patients when compared with LADA, the level of another potent pro-inflammatory cytokine, namely IL-18, remained unchanged between participants from both groups. The current positive correlation between IL-18 and IL-6, together with hs-CRP, is in line with a previously observed association between them and the metabolic syndrome (22). The latter was much more prevalent in T2D even in BMI-matched participants in the current study. Although correlations of IL-18 with Hb1Ac, PPG and HDL cholesterol have been previously described in T1D (55), here they were not confirmed in subjects with LADA. One reason for the lack of association between lipid, glycemic and

<table>
<thead>
<tr>
<th>Table 2 Correlation coefficients between IL-18 and clinical, metabolic and inflammatory parameters.</th>
<th>T2D (n=76)</th>
<th>LADA (n=24)</th>
<th>Controls (n=14)</th>
<th>T2D (n=28)*</th>
<th>LADA (n=20)*</th>
</tr>
</thead>
<tbody>
<tr>
<td>BMI (kg/m²)</td>
<td>0.015</td>
<td>0.897</td>
<td>0.210</td>
<td>0.325</td>
<td>0.004</td>
</tr>
<tr>
<td>WC (cm)</td>
<td>0.073</td>
<td>0.532</td>
<td>0.297</td>
<td>0.169</td>
<td>0.064</td>
</tr>
<tr>
<td>SBP (mmHg)</td>
<td>−0.037</td>
<td>0.749</td>
<td>0.289</td>
<td>0.171</td>
<td>0.527</td>
</tr>
<tr>
<td>DBP (mmHg)</td>
<td>−0.094</td>
<td>0.418</td>
<td>0.133</td>
<td>0.535</td>
<td>0.507</td>
</tr>
<tr>
<td>FPG (mmol/L)</td>
<td>0.075</td>
<td>0.530</td>
<td>−0.098</td>
<td>0.655</td>
<td>0.483</td>
</tr>
<tr>
<td>PPG (mmol/L)</td>
<td>0.244</td>
<td>0.043</td>
<td>0.027</td>
<td>0.907</td>
<td>–</td>
</tr>
<tr>
<td>HbA1c (%)</td>
<td>0.007</td>
<td>0.956</td>
<td>0.168</td>
<td>0.434</td>
<td>–</td>
</tr>
<tr>
<td>LDL-C (mmol/L)</td>
<td>−0.141</td>
<td>0.251</td>
<td>0.134</td>
<td>0.563</td>
<td>−0.500</td>
</tr>
<tr>
<td>HDL-C (mmol/L)</td>
<td>−0.377</td>
<td>0.001</td>
<td>−0.392</td>
<td>0.079</td>
<td>0.500</td>
</tr>
<tr>
<td>TG (mmol/L)</td>
<td>0.072</td>
<td>0.546</td>
<td>0.301</td>
<td>0.154</td>
<td>−0.172</td>
</tr>
<tr>
<td>hs-CRP (mg/L)</td>
<td>0.427</td>
<td>0.001</td>
<td>0.113</td>
<td>0.832</td>
<td>−0.241</td>
</tr>
<tr>
<td>IL-6 (pg/mL)</td>
<td>0.398</td>
<td>&lt;0.001</td>
<td>−0.131</td>
<td>0.541</td>
<td>−0.241</td>
</tr>
</tbody>
</table>

*Correlations within groups with BMI-matched participants.

BMI, body mass index; DBP, diastolic blood pressure; FPG, fasting plasma glucose; HDL-C, high density lipoprotein cholesterol; Hs-CRP, high-sensitivity C-reactive protein; IL-6, interleukin 6; IL-18, interleukin-18; LADA, latent autoimmune diabetes of the adults; LDL-C, low density lipoprotein cholesterol; PPG, postprandial plasma glucose; SBP, systolic blood pressure; T2D, type 2 diabetes; TG, triglycerides; WC, waist circumference.
inflammatory parameters in these participants could be the lower prevalence of the metabolic syndrome, as the association of IL-18 with the latter seems to be stronger than that with obesity (22).

Another possible explanation concerns a dominant role of IL-18 in the autoimmune pathogenesis of LADA and not the insulin resistance-associated chronic low-grade inflammation in this type of diabetes mellitus. Although beta-cell dysfunction in T2D has recently also been associated with insulitis (56), it is currently not clear whether IL-18 contributes to beta-cell apoptosis in T2D as has been demonstrated for other cytokines (57).

Lack of association between IL-18 and the described metabolic and inflammatory parameters could even result from both the metabolic and immunologic factors discussed. It has been shown that LADA could present with a heterogeneous clinical and immunologic characteristics moving it closer to either T1D or T2D (58). A larger cohort of participants with LADA with both phenotypes and related GAD65A titer (59) would clarify the role of IL-18 in metabolic control-associated low-grade chronic inflammation and autoimmune process. The main limitations of the study are the relatively low number of participants, a higher percentage of the female gender in all groups and the significant difference in the prevalence of the metabolic syndrome between groups, even when they were BMI matched. However, we have not been able to find another study that compares the IL-18 serum level between T2D and LADA, and these results add to current knowledge of the cytokine’s place in diabetes mellitus and prompts further investigations of the subject.

Conclusion
The current study showed a higher serum level of IL-18 in patients with T2D and LADA than in the control subjects, but a similar one between both diabetic groups even after matching them for BMI and glycemic control. Correlations with glycemic, lipid and inflammatory parameters were observed in T2D only. Further analysis is needed to distinguish the role of IL-18 in insulin-resistance-associated chronic low-grade inflammation from that in the autoimmune response in LADA.

Declaration of interest
The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

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Acknowledgements
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